

ASSESSING REAL-TIME SICKLING KINETICS IN INDIVIDUAL CELLS FROM ERYTHROCYTE POPULATIONS OF SUBJECTS WITH DIFFERENT HbA, HbS AND HbF PERCENTAGES FUNCTIONAL FLUIDICS ALIYA U. ZAIDI, PHD¹, XIUFENG GAO, MD, MSC¹, ROBERT GOODRICH, BSC¹, MARTA FERRANTI, MSC¹, RASA BORHAN, BSC¹, KE LIU, PHD^{1*}, NNAMDI OKEKE, PHARMD¹, MICHAEL TARASEV, PHD, MBA¹, MARIAMA KABORE², COURTNEY D. FITZHUGH, MD², PRIYA S. CHOCKALINGAM, PHD³ AND PATRICK C. HINES, MD, PHD^{1,4}. ¹Functional Fluidics Inc., Detroit, MI²NHLBI/NIH, Bethesda, MD³Beam Therapeutics, Cambridge, MA⁴Wayne State University Division of Pediatric Critical Care Medicine, Detroit, MI

INTRODUCTION

- Polymerization of deoxygenated hemoglobin S (HbS) leads to red blood cell (RBC) sickling and vaso-occlusion
- Allogeneic hematopoietic stem cell transplantation (HSČT) is the only approved curative option available for sickle cell disease (SCD)
- Objective biomarkers are needed to assess disease status, population-scale sickling kinetics and to monitor the pharmacodynamic impact of Hbmodifying and curative therapies
- Functional Fluidics has developed a dynamic sickling assay (DSA) that uses an enzymatic O_2 scavenging system providing tight control over the rate and depth of induced-hypoxia

AIM

Assess real-time RBC sickling kinetics in sickle cell trait (SCT) and SCD patients with variable distributions of HbA, HbS and HbF, and compare pre-and post-HSCT

METHOD

Dynamic Sickling Assay (Figure 1)

- Hypoxia is induced through a timed protocatechuic acid/protocatechuate-3,4dioxygenase enzymatic reaction (PCD-PCA)
- The fraction of sickled cells is determined by timelapse imaging, analyzed with AI technology and reported as a function of time
- Measured parameters include:
- mPoS@5%: time to reach 5% induced sickling (min)
- mPoS@50%: time to reach 50% induced sickling (min)
- **Rate of Sickling**: highest induced sickling rate $(\% \cdot \text{minute}^{-1})$
- Maximum Sickling: highest sickling percentage
- **AUC at 10min**: The area under the curve during 10min of DSA (%·minute)

Sample collection

- SCT and SCD subjects, with and without chronic exchange transfusions were enrolled in The Institute for Regenerative and Cellular Medicine (IRCM) IRB under the RBC health initiative protocol (FF-RBC-003v5)
- SCD subjects were enrolled in NIH haploidentical HSCT protocol 17-H-0069. Blood samples from 4/10 planned subjects were collected pre-and 3, 6, and 12 months after HSCT





variable HbF (**C**)

CONCLUSIONS

SCT subjects with similar HbS% exhibit different sickling kinetics upon exposure to the same hypoxic stress due to intracellular RBC distribution of HbA and HbS

Transfusion SCD samples show an improvement in sickling kinetics as HbS% decreases approaching a significant correlation with AUC10

Statistically significant correlations are observed with increasing HbF percentages on 4 out of 5 sickling parameters

 Compared to pre-transplant values, post-HSCT samples exhibit significant improvement in: mPOS@5%, rate of sickling, maximum sickling and AUC10

DSA can differentiate samples based on Hb distribution and can be used to assess sickling kinetics pre- and post HbS modifying and curative therapies, such as hematopoietic stem cell (HSCT)

Representative DSA Profile 100 80 60 20

Figure 3. Sickling kinetics in SCD subjects pre-and post HSCT (A) Representative sickling profile from 1 patient (B) Comparative analyses from 4 SCD subjects *only 1 post-HSCT sample exhibits >50% max. sickling

Time, minutes

Table 1. Sickling kinetics correlations

		<u>mPoS@5%</u>		<u>mPoS@50%</u>		Rate of Sickling		Max_Sickling		<u>AUC10</u>	
	n	r	р	r	р	r	р	r	р	r	р
HbS% (Sickle Cell Trait)	3	NA	NA	NA	NA	NA	NA	0.30	0.803	0.86	0.345
HbS% (Transfused SCD)	3	-0.99	0.100	NA	NA	0.626	0.596	0.99	0.107	0.99	0.066
HbF% (non-Transfused SCD)	9	0.87	0.002	0.83	0.006	-0.74	0.023	0.15	0.692	0.73	0.025
 No significant correlation conditions in samples with HbS% and DSA paramete likely due to limited sample 	in SC 1 sim rs sh e siz	CT subj ilar Hb iow a d ze	ects, c S% lev correlc	althoug els ation ir	gh note n SCD t	eworthy ransfuse	differer ed samp	nces ev les, no	vident u t statist	under h ically s	nypoxie

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RESULTS

POST-HSCT SAMPLES EXHIBIT IMPROVED SICKLING KINETICS



Compared to baseline values, post-HSCT samples exhibit statistically significant improvement in: mPOS@5%, rate of sickling, maximum sickling and AUC10

HbF% demonstrates a significantly strong correlation with DSA parameters except for max. sickling

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CONTACT INFORMATION

